

Amino Acid Nutrition During the Transition Period for Dairy Cows

F.C. Cardoso¹

*Department of Animal Sciences
University of Illinois*

Introduction

During the transition period from late gestation through early lactation, the dairy cow undergoes tremendous metabolic adaptations (Bell, 1995). The endocrine changes during the transition period are necessary to prepare the dairy cow for parturition and lactogenesis. As peak milk yield increases, the transition period for dairy cows becomes much more challenging with most infectious diseases and metabolic disorders occurring during this time (Grummer, 1995; Drackley, 1999). Decreased dry matter intake (**DMI**) during late gestation influences metabolism, leading to fat mobilization from adipose tissue and glycogen from liver.

Nutrient demand for milk synthesis is increased in early lactation; if no compensatory intake of nutrients is achieved to cope with the requirement, reproductive functions (i.e., synthesis and secretion of hormones, follicle ovulation, and embryo development) may be depressed. Milk production increases faster than energy intake in the first 4 to 6 weeks after calving, and thus, high yielding cows will experience negative energy balance (**NEB**). Nutritional strategies and feeding management during pre-calving and post-calving periods impact health, productivity, and fertility of high producing dairy cows. Formulating diets to meet requirements of the cows but avoid over-consumption of energy may improve outcomes

of the transition period and lead to improved fertility. Management to improve cow comfort and ensure good intake of the ration is pivotal for success. Impacts of the transition program should be evaluated in a holistic way that considers disease occurrence, productivity, and fertility.

Studies over the last 2 decades clearly established the link between nutrition and fertility in ruminants (Robinson et al., 2006; Wiltbank et al., 2006; Grummer et al., 2010; Santos et al., 2010; Cardoso et al., 2013; Drackley and Cardoso, 2014). Dietary changes can cause an immediate and rapid alteration in a range of humoral factors that can alter endocrine and metabolic signaling pathways crucial for reproductive function (Boland et al., 2001; Diskin et al., 2003). Moreover, periconceptional nutritional environment in humans and other animals is critical for the long-term setting of postnatal phenotype (Fleming et al., 2015). Restricting the supply of B-vitamins and methionine during the periconceptional period in sheep, e.g., resulted in adverse cardiometabolic health in postnatal offspring (Sinclair et al., 2007). Feeding female mice a low-protein diet during the preimplantation period of pregnancy resulted in a reduction in amino acid (**AA**) concentration in uterine fluid and serum and attendant changes in the AA profile of the blastocyst (Eckert et al., 2012).

¹Contact at: 290 Animal Sciences Laboratory, 1207 West Gregory Dr., Urbana, IL 61801, (217) 300-2303, Email: cardoso2@illinois.edu.



Strategies have been used to improve the reproductive performance of dairy cows through alteration of nutritional status (Santos et al., 2001; Santos et al., 2008). In other species, dietary supplementation with specific AA (e.g., arginine, glutamine, leucine, glycine, and methionine) had beneficial effects on embryonic and fetal survival and growth through regulation of key signaling and metabolic pathways (Wang et al., 2012; Del Curto et al., 2013). Methionine and lysine are the most limiting AA in lactating cows (NRC, 2001), but supplementation of diets with crystalline methionine and lysine has been excluded because free methionine and lysine are quickly and almost totally degraded by the microorganisms in the rumen (NRC, 2001). In contrast, supplementing rumen-protected methionine (**RPM**) and rumen-protected lysine (**RPL**) has a positive effect on milk protein synthesis in dairy cows (Pisulewski et al., 1996; NRC, 2001; Ordway, et al., 2009; Osorio et al., 2013). Although the role of methionine in bovine embryonic development is unknown, there is evidence that methionine availability alters the follicular dynamics of the first dominant follicle (Acosta et al., 2017), the transcriptome of bovine preimplantation embryos in vivo (Penagaricano et al., 2013), and its contents (Acosta et al., 2016).

Reproduction, Nutrition, and Health

A widespread assumption is that fertility of modern dairy cows is decreasing, particularly for Holstein-Friesen genetics, at least in part because of unintended consequences of continued selection for high milk production. This assumption has been challenged recently (LeBlanc, 2010; Bello et al., 2012). There is a wide distribution of reproductive success both within and among herds. For example, within 5 California herds encompassing 6,396 cows, cows in the lowest quartile for milk yield in the first 90 days postpartum (32.1 kg/day) were less

likely to have resumed estrous cycles by 65 days postpartum than cows in quartiles two (39.1 kg/day), three (43.6 kg/day), or four (50.0 kg/day); milk production did not affect risk for pregnancy (Santos et al., 2009). Changes in management systems and inadequacies in management may be more limiting for fertility of modern dairy cows than their genetics per se.

Dairy cows are susceptible to production disorders and diseases during the periparturition period and early lactation, including milk fever, ketosis, fatty liver, retained placenta, displaced abomasum, metritis, mastitis, and lameness (Mulligan et al., 2006; Ingvarsen and Moyes, 2013; Roche et al., 2013). There is little evidence that milk yield per se contributes to greater disease occurrence. However, peak disease incidence (shortly after parturition) corresponds with the time of greatest NEB, the peak in blood concentrations of nonesterified fatty acids (**NEFA**), and the greatest acceleration of milk yield (Ingvarsen et al., 2003). Peak milk yield occurs several weeks later. Disorders associated with postpartum NEB also are related to impaired reproductive performance, including fatty liver (Rukkwamsuk et al., 1999; Jorritsma et al., 2003) and ketosis (Walsh et al., 2007; McArt et al., 2012). Cows that lost >1 body condition score (BCS) unit (1-5 scale) had greater incidence of metritis, retained placenta, and metabolic disorders (displaced abomasum, milk fever, and ketosis), as well as a longer interval to first breeding than cows that lost <1 BCS unit during the transition (Kim and Suh, 2003).

Indicators of NEB are highly correlated with lost milk production, increased disease, and decreased fertility (Ospina et al., 2010; Chapinal et al., 2012). However, the extent to which NEB is causative for periparturition health problems rather than just a correlated phenomenon must be examined critically (Roche et al., 2013).

For example, in transition cows, inflammatory responses may decrease DMI, cause alterations in metabolism, and predispose cows to greater NEB or increased disease (Bertoni et al., 2008; Graugnard et al., 2012 and 2013; Ingvarlsen and Moyes, 2013). Inducing a degree of calculated NEB in mid-lactation cows similar to what periparturient cows often encounter does not result in marked increases in ketogenesis or other processes associated with periparturient disease (Moyes et al., 2009). Nevertheless, early postpartal increases in NEFA and decreases in glucose concentrations were strongly associated with pregnancy at first insemination in a timed artificial insemination (TAI) program (Garverick et al., 2013). Although concentrations of NEFA and glucose were not different between cows that ovulated or did not before TAI, probability of pregnancy decreased with greater NEFA and increased with greater glucose concentrations at day 3 postpartum (Garverick et al., 2013). In support of these findings, early occurrence of subclinical ketosis is more likely to decrease milk yield and compromise fertility. McArt et al. (2012) reported that cows with subclinical ketosis detected between 3 to 7 days after calving were 0.7 times as likely to conceive to first service and 4.5 times more likely to be removed from the herd within the first 30 days in milk compared with cows that developed ketosis at 8 days or later.

Cows that successfully adapt to lactation (Jorritsma et al., 2003) and can avoid metabolic (Ingvarlsen et al., 2003) or physiological imbalance (Ingvarlsen and Moyes, 2013) are able to support both high milk production and successful reproduction while remaining healthy. Decreased fertility in the face of increasing milk production may be attributable to greater severity of postpartal NEB resulting from inadequate transition management or increased rates of disease. Competition for nutrients between the divergent outcomes of early

lactation and subsequent pregnancy will delay reproductive function. Because NEB interrupts reproduction in most species, including humans, inappropriate nutritional management may predispose cows to both metabolic disturbances and impaired reproduction. Cows must make “metabolic decisions” about where to direct scarce resources, and in early lactation, nutrients will be directed to milk production rather than to the next pregnancy (Friggens, 2003).

Different nutritional strategies have been proposed to improve reproduction of the dairy cow with no detrimental effect on lactation performance. Feeding high quality forages, controlled-energy (CE) diets, or adding supplemental fat to diets are some of the most common ways to improve energy intake in cows (Cardoso et al., 2013; Drackley and Cardoso, 2014; Mann et al., 2015). Reproduction of dairy cattle may be benefited by maximizing DMI during the transition period, minimizing the incidence of periparturient problems (Cardoso et al., 2013; Drackley and Cardoso, 2014).

Prepartum Dietary Considerations

Controlling energy intake during the dry period to near calculated requirements leads to better transition success (Grum et al., 1996; Dann et al., 2005 and 2006; Douglas et al., 2006; Janovick et al., 2011; Graugnard et al., 2012 and 2013; Ji et al., 2012). Research drew from earlier reports that limiting nutrient intakes to requirements of the cows was preferable to over-consumption of energy (e.g., Kunz et al., 1985). Cows fed even moderate-energy diets (1.50 to 1.60 Mcal NEL/kg DM) will easily consume 40 to 80% more NEL than required during both far-off and close-up periods (Dann et al., 2005 and 2006; Douglas et al., 2006; Janovick and Drackley, 2010). Cows in these studies were all less than 3.5 BCS (1-5 scale) at dry-off and were fed individually TMR based

on corn silage, alfalfa silage, and alfalfa hay with some concentrate supplementation. We have no evidence that the extra energy and nutrient intake was beneficial in any way. More importantly, our data indicate that allowing cows to over-consume energy even to this degree may predispose them to health problems during the transition period if they face stressors or challenges that limit DMI (Cardoso et al., 2013).

Prolonged over-consumption of energy during the dry period can decrease post-calving DMI (Dann et al., 2006; Douglas et al., 2006; Janovick and Drackley, 2010). Over-consuming energy results in negative responses of metabolic indicators, such as higher NEFA and betahydroxybutyrate (**BHB**) in blood and more triacylglycerol (**TAG**) in the liver after calving (Douglas et al., 2006; Janovick et al., 2011). Alterations in cellular and gene-level responses in liver (Lor et al., 2006 and 2007) and adipose tissue (Ji et al., 2012) potentially explain many of the changes at the cow level. Over-consumption of energy during the close-up period increases the enzymatic “machinery” in adipose tissue for TAG mobilization after calving, with transcriptional changes leading to decreased lipogenesis, increased lipolysis, and decreased ability of insulin to inhibit lipolysis (Ji et al., 2012). Controlling energy intake during the dry period also improved neutrophil function postpartum (Graugnard et al., 2012) and so may lead to better immune function.

Allowing dry cows to consume more energy than required, even if cows do not become noticeably over-conditioned, results in responses that would be typical of overly fat cows. Because energy that cows consume in excess of their requirements must either be dissipated as heat or stored as fat, we speculated that the excess is accumulated preferentially in internal adipose tissue depots in some cows. Moderate over-consumption of energy by non-lactating cows

for 57 days led to greater deposition of fat in abdominal adipose tissues (omental, mesenteric, and perirenal) than in cows fed a high-bulk diet to control energy intake to near requirements (Drackley et al., 2014). The NEFA and signaling molecules released by visceral adipose tissues travel directly to the liver, which may cause fatty liver, subclinical ketosis, and secondary problems with liver function.

Data from our studies support field observations that controlled-energy dry cow programs decrease health problems (Beever, 2006). Other research groups (Rukkwamsuk et al., 1998; Holcomb et al., 2001; Holtenius et al., 2003; Vickers et al., 2013) have reached similar conclusions about controlling energy intake during the dry period, although not all studies have shown benefits (Winkleman et al., 2008). Application of these principles can be through controlled limit-feeding of moderate energy diets or ad libitum feeding of high-bulk, low-energy rations (Janovick and Drackley, 2010; Janovick et al., 2011; Ji et al., 2012) as proposed by others (Beever, 2006).

Nutritionally complete diets must be fed and the TMR must be processed appropriately so that cows do not sort the bulkier ingredients (Janovick and Drackley, 2010). Feeding bulky forage separately from a partial TMR or improper forage processing will lead to variable intake among cows, with some consuming too much energy and some too little. Underfeeding relative to requirements, where nutrient balance also is likely limiting, leads to increased incidence of retained placenta and metritis (Mulligan et al., 2006). Merely adding a quantity of straw to a diet is not the key principle; rather, the diet must be formulated to limit the intake of energy (approximately 1.3 Mcal NE_L/kg DM, to limit intake to about 15 Mcal/day for typical Holstein cows) but meet the requirements for protein, minerals, and vitamins. Reports of

increased transition health problems or poor reproductive success (Whitaker et al., 1993) with “low energy” dry cow diets must be examined carefully to discern whether nutrient intakes were adequate.

Fresh Cow (postpartum) Dietary Considerations

Less is known about diet formulation for the immediate postpartum period to optimize transition success and subsequent reproduction. Increased research is needed in this area. Proper dietary formulation during the dry period or close-up period will maintain or enable rumen adaptation to higher grain diets after calving. Failure to do so may compromise early lactation productivity. For example, Silva-del-Rio et al. (2010) attempted to duplicate the dietary strategy of Dann et al. (2006) by feeding either a low-energy far-off diet for 5 weeks followed by a higher-energy diet for the last 3 weeks before parturition, or by feeding the higher-energy diet for the entire 8-week dry period. They found that cows fed the higher-energy diet for only 3 weeks before parturition produced less milk than cows fed the diet for 8 weeks (43.8 vs. 48.5 kg/day). However, the far-off dry period diet contained 55.1% alfalfa silage and 38.5% wheat straw but no corn silage. In comparison, the higher-energy dry period diet and the early lactation diet both contained 35% corn silage. Ruminal adaptation likely was insufficient for cows fed the higher energy diet for only 3 weeks.

A major area of concern in the fresh cow period is a sudden increase in dietary energy density, leading to subacute ruminal acidosis (**SARA**), which can decrease DMI and digestibility of nutrients (Mulligan and Doherty, 2008). Adequate physical form of the diet, derived either from ingredients or mixing strategy, must be present to stimulate ruminal activity and chewing behavior (Zabeli and

Metzler-Zabeli, 2012), although good methods to quantify “adequacy” remain elusive. Dietary starch content and fermentability likely interact with forage characteristics and ration physical form. Dann and Nelson (2011) compared three dietary starch contents (primarily from corn starch) in the fresh cow period for cows fed a controlled energy-type ration in the dry period. Milk production was greatest when starch content was moderate (23.2% of DM) or low (21.0% of DM) in the fresh cow diet compared with high (25.5% of DM). If SARA decreases DMI and nutrient availability to the cow, NEFA mobilization and increased ketogenesis may follow. In addition, rapid starch fermentation in the presence of NEFA mobilization leads to bursts of propionate reaching the liver, which may decrease feeding activity and DMI according to the hepatic oxidation theory (Allen et al., 2009). A moderate starch content (ca. 23 to 25% of DM) with starch of moderate fermentability (for example, ground dry corn rather than high-moisture corn or ground barley) along with adequate effective forage fiber may be the best strategy for fresh cows. Recent research also has demonstrated that high grain diets can lead to greater numbers of gram-negative bacteria such as *E. coli* with resulting increases in endotoxin present in the rumen, which may decrease barrier function and inflammatory responses in the cow (Zabeli and Metzler-Zabeli, 2012).

Supplemental fats have been widely investigated as a way to increase dietary energy intake and improve reproduction (Thatcher et al., 2011). A novel strategy to use polyunsaturated fatty acid (**PUFA**) supplements to improve reproduction has been reported (Silvestre et al., 2011). Cows fed calcium salts of safflower oil from 30 days before to 30 days after calving, followed by calcium salts of fish oil to 160 days postpartum, had greater pregnancy rates and higher milk production. The mechanism

is believed to be provision of greater amounts of linoleic acid (omega-6 PUFA) until early postpartum, which improves uterine health, followed by greater amounts of omega-3 PUFA from fish oil to decrease early embryonic loss (Thatcher et al., 2011). The effects of turbulent transitions on reproduction are established early postpartum, likely during the first 10 days to 2 weeks postpartum (Butler, 2003; McArt et al., 2012; Garverick et al., 2013). By 8 weeks postpartum, >95% of cows should be at or above energy balance (Sutter and Beever, 2000). Use of targeted prepartum and postpartum strategies may minimize health problems and lessen NEB, and thereby improve subsequent fertility.

Body Condition Score

The role of excessive BCS in contributing to transition problems and impaired subsequent reproduction is well established and has been discussed by many authors (Drackley et al., 2005; Garnsworthy et al., 2008; Roche et al., 2013). Cows with excessive body lipid reserves mobilize more of that lipid around calving, have poorer appetites and DMI before and after calving, have impaired immune function, have increased indicators of inflammation in blood and may be more subjected to oxidative stress (Contreras and Sordillo, 2011). What constitutes “excessive” BCS relative to the cow’s biological target remains controversial. Garnsworthy (2007) argued that the average optimal BCS has decreased over time with increased genetic selection for milk yield, perhaps related to correlated changes in body protein metabolism. Recommendations for optimal BCS at calving have trended downward over the last two decades, and in the author’s opinion, a score of about 3.0 (1-5 scale) represents a good goal at present. Adjustment of average BCS should be a longstanding project and should not be undertaken during the dry period.

Cows fed high-energy (1.58 Mcal NE_L/kg DM) diets during the last 4 weeks before calving lost more BCS in the first 6 weeks postpartum than those fed controlled energy (1.32 Mcal NE_L/kg DM) diets (−0.43 and −0.30, respectively) (Cardoso et al., 2013). The effect of BCS change on the cow’s fertility is clear. Carvalho et al. (2014) showed that cows that either gained or maintained BCS from calving to 21 days after calving had higher (38.2 and 83.5%, respectively) pregnancy per AI at 40 days than cows that lost BCS (25.1%) during that same period. Previously, Santos et al. (2009) had shown that cows that had > 1.0 BCS unit change from calving to AI at approximately 70 days postpartum had lower pregnancy per AI (28%) than cows that lost < 1.0 BCS unit change (37.3%) or did not have a BCS change (41.6%). In a grazing system, researchers from New Zealand suggested that BCS at calving should be targeted at 2.75 to 3.0 to optimize production, while reducing liver lipid accumulation and the negative effects of inflammation on liver function (Roche et al., 2013; Akbar et al., 2015).

The Importance of Amino Acids

Some AA are limiting for optimal milk production as evidenced by an increase in milk yield, percentage of milk protein, and milk protein yield after supplementation with specific, rumen-protected amino acids. The first 3 limiting amino acids for milk production are considered to be methionine, lysine (NRC, 2001), and histidine (Hutannen, et al., 2002). In addition, many amino acids can have positive effects on physiological processes that are independent of their effects on synthesis of proteins (Wu, et al., 2013). Fertilization and the first few days of embryo development occur in the oviduct. By about 5 days after estrus, the embryo arrives in the uterine horn. The embryo reaches the blastocyst stage by 6 to 7 days after estrus. The embryo hatches from the zona pellucida by

about day 9 after estrus and then elongates on days 14 to 19. The elongating embryo secretes the protein interferon-tau that is essential for rescue of the corpus luteum and continuation of the pregnancy. By days 25 to 28, the embryo attaches to the caruncles of the uterus and begins to establish a vascular relationship with the dam through the placenta. During all the time prior to embryo attachment, the embryo is free-floating and is dependent upon uterine secretions for energy and the building blocks for development, including amino acids. Thus, it is critical to understand the changes in amino acid concentrations in the uterus that accompany these different stages of embryo development.

The lipid profile of oocytes and early embryos can be influenced by the environment of the cow. Our group ran a trial with the objective to determine the effect of supplementing rumen-protected methionine on DNA methylation and lipid accumulation in preimplantation embryos of dairy cows (Acosta et al., 2016). Lactating Holstein cows entering their 2nd or greater lactation were randomly assigned to 2 treatments from 30 ± 2 DIM to 72 ± 2 DIM; Control (CON; n = 5, fed a basal diet with a 3.4:1 Lys:Met) and Methionine (MET; n = 5, fed the basal diet plus Smartamine M to a 2.9:1 Lys:Met). Embryos were flushed 6.5 days after artificial insemination. Embryos with stage of development 4 or greater were used for analysis. For lipids, fluorescence intensity of Nile Red staining was compared against a negative control embryo (subtraction of background). A total of 37 embryos were harvested from cows (MET = 16; CON = 21). Cows receiving MET had greater lipid accumulation (7.3 arbitrary units) when compared with cows receiving CON (3.7 arbitrary units). There were no treatment effects on number of cells or stage of development. In conclusion, cows supplemented with methionine produced embryos with higher lipid concentration when compared to CON,

which could potentially serve as an important source of energy for the early developing embryo.

The requirements for complete development of bovine embryos have not yet been determined. Current culture conditions allow development of bovine embryos to the blastocyst stage (day 7 to 8) and even allow hatching of a percentage of embryos (day 9); however, conditions have not been developed in vitro that allow elongation of embryos. The methionine requirements for cultured pre-implantation bovine embryos (day 7 to 8) was determined in studies from University of Florida (Bonilla et al., 2010). There was a surprisingly low methionine requirement ($7 \mu\text{M}$) for development of embryos to the blastocyst stage by day 7; however, development to the advanced blastocyst stage by day 7 appeared to be optimized at around $21 \mu\text{M}$ (Bonilla et al., 2010). Thus, the results of these studies indicated that development of morphologically normal bovine embryos did not require elevated methionine concentrations ($>21 \mu\text{M}$), at least during the first week after fertilization. Stella (2017) reported the plasma concentration of cows fed RPM or not (CON) (Figure 1). It seems that cows when fed RPM have plasma methionine concentration greater than $20 \mu\text{M}$.

Researchers at the Univ. of Wisconsin (Toledo et al., 2015) conducted a trial with a total of 309 cows (138 primiparous and 171 multiparous) that were blocked by parity and randomly assigned to 2 treatments: 1) CON: Cows fed a ration formulated to deliver 2500 g of metabolizable protein (MP) with 6.9% Lys (% of MP) and 1.9 Met (% of MP), and 2) RPM: Cows fed a ration formulated to deliver 2500 g of MP with 6.9% Lys, % of MP) and 2.3 % Met (% of MP). Cows were randomly assigned to 3 pens with head-locks and fed a single basal TMR twice daily. From 28 to 128 DIM, after the AM

milking, cows were head-locked for 30 minutes and the TMR of CON and RPM cows were individually top dressed with 50 g of DDG or 50 g of a mix of DDG (29 g) and Smartamine M (21 g), respectively. Following a double ovsynch protocol, cows were inseminated and pregnancy checked at 28 (plasma Pregnancy Specific Protein-B concentration), 32, 47, and 61 days (ultrasound). Individual milk samples were taken once a month and analyzed for composition. There were no statistical differences in milk production, but RPM cows had a higher milk protein concentration. Cows fed the methionine enriched diet had a lower pregnancy loss from 21 to 61 after AI (16.7% RPM cows vs. 10.0% from CON cows). Pregnancy losses between days 28 and 61 were not different in the primiparous cows (12.8% CON and 14.6% RPM), however, pregnancy losses between treatments were significant for the multiparous cows (19.6% CON vs. 6.1% RPM; Toledo et al., 2015).

Perhaps the most detrimental impact of NEB on reproductive performance is delayed return to cyclicity (Jorritsma et al., 2003). The dominant follicle (DF) growth and estradiol (E_2) production are key factors for a successful conception, and their impairment can be attributed to reduced luteinizing hormone (LH) pulses (Grainger et al., 1982), as well as decreased circulating insulin and IGF-I concentrations (Canfield and Butler, 1990; Komaragiri and Erdman, 1997). Furthermore, immune function is also suppressed along the periparturient period (Butler 2003; Kehrlı et al., 1999); NEB and fatty liver syndrome were demonstrated to impair peripheral blood neutrophil function (Zerbe et al., 2000; Hammon et al., 2006). Acosta et al. (2017) reported that methionine and choline supplementation induced a down regulation of pro-inflammatory genes, possibly indicating lower inflammatory processes in follicular cells of the first DF postpartum. Also, supplementing methionine, during the

transition period increased 3β -hydroxysteroid dehydrogenase expression in the follicular cells of the first DF postpartum. It is important to highlight that higher methionine concentrations in the follicular fluid of supplemented cows can potentially affect oocyte quality. The understanding on how this finding may affect reproductive performance in commercial farms needs to be further investigated. Batistel et al. (2017) reported that that studies with non-ruminant species argue for the potential relevance of the maternal methionine supply during late gestation in enhancing utero-placental uptake and transport of nutrients. The authors hypothesized that the greater newborn body weight from cows fed RPM compared to control (42 vs. 44 kg) could have been a direct response to the greater nutrient supply from the feed intake response induced by methionine, the fact that certain amino acids and glucose are known to induce mTOR signaling to different degrees is highly suggestive of “nutrient specific” mechanistic responses.

Conclusions

Formulation and delivery of appropriate diets that limit total energy intake to requirements but also provide proper intakes of all other nutrients before calving can help lessen the extent of NEB after calving. Effects of such diets on indicators of metabolic health are generally positive, suggesting the potential to lessen effects of periparturient disease on fertility. Supplementation of cows with methionine during the final stages of follicular development and early embryo development, until day 7 after breeding, lead to lipid accumulation changes in the embryos and resulted in differences in gene expression in the embryo. Methionine supplementation seems to impact the preimplantation embryo in a way that enhances its capacity for survival because there is strong evidence that endogenous lipid reserves serve

as an energy substrate. The lower pregnancy losses from cows fed a methionine enriched diet suggest that methionine favors the embryo survival, at least in multiparous cows.

References

- Acosta, D.A.V., A.C. Denicol, P. Tribulo, M.I. Rivelli, C. Skenandore, Z. Zhou, D. Luchini, M.N. Corrêa, P.J. Hansen, and F.C. Cardoso. 2016. Effects of rumen-protected methionine and choline supplementation on the preimplantation embryo in Holstein cows. *Theriogenology* 85:1669-1679.
- Acosta, D.A.V., M.I. Rivelli, C. Skenandore, Z. Zhou, D.H. Keisler, D. Luchini, M.N. Corrêa, and F.C. Cardoso. 2017. Effects of rumen-protected methionine and choline supplementation on steroidogenic potential of the first postpartum dominant follicle and expression of immune mediators in Holstein cows. *Theriogenology* 96:1-9.
- Akbar, H., T.M. Grala, M. Vailati Riboni, F.C. Cardoso, G. Verkerk, J. McGowan, K. Macdonald, J. Webster, K. Schutz, S. Meier, L. Matthews, J.R. Roche, and J.J. Loor. 2015. Body condition score at calving affects systemic and hepatic transcriptome indicators of inflammation and nutrient metabolism in grazing dairy cows. *J. Dairy Sci.* 98:1019–1032.
- Allen, M.S., B.J. Bradford, and M. Oba. 2009. Board Invited Review: The hepatic oxidation theory of the control of feed intake and its application to ruminants. *J. Anim. Sci.* 87:3317-3334.
- Batistel, F., A.S.M. Alharthi, L. Wang, C. Parys, Y. Pan, F.C. Cardoso, and J.J. Loor. 2017. Placentome nutrient transporters and mTOR signaling proteins are altered by methionine supply during late-gestation in dairy cows and are associated with newborn birth weight. *The Journal of Nutrition.* 147:1640-1647.
- Beever, D.E. 2006. The impact of controlled nutrition during the dry period on dairy cow health, fertility and performance. *Animal Reproduction Science* 96:212-226.
- Bell, A.W. 1995. Regulation of organic nutrient metabolism during transition from late pregnancy to early lactation. *J. Anim. Sci.* 73:2804-2819.
- Bello, N.M., J.S. Stevenson, and R.J. Tempelman. 2012. Invited review: Milk production and reproductive performance: Modern interdisciplinary insights into an enduring axiom. *J. Dairy Sci.* 95:5461-5475.
- Bertoni, G., E. Trevisi, X. Han, and M. Bionaz. 2008. Effects of inflammatory conditions on liver activity in puerperium period and consequences for performance in dairy cows. *J. Dairy Sci.* 91:3300-3310.
- Boland, M.P., P. Lonergan, and D. O’Callaghan. 2001. Effect of nutrition on endocrine parameters, ovarian physiology, and oocyte and embryo development. *Theriogenology.* 55:1323–1340.
- Bonilla, L,D. Luchini, E. Devillard, and P.J. Hansen. 2010. Methionine requirements for the preimplantation bovine embryo. *J. Reprod. Dev.* 56:527–532.
- Butler, W.R. 2003. Energy balance relationships with follicular development, ovulation and fertility in postpartum dairy cows. *Livestock Prod. Sci.* 83:211-218.



- Canfield R.W., and W.R. Butler. 1990. Energy balance and pulsatile LH secretion in early postpartum dairy cattle. *Domest. Anim. Endocrinol.* 7:323-330.
- Cardoso, F.C., S.J. LeBlanc, M.R. Murphy, and J.K. Drackley. 2013. Prepartum nutritional strategy affects reproductive performance in dairy cows. *J. Dairy Sci.* 96:5859–5871.
- Carvalho, P.D., A.H. Souza, M.C. Amundson, K.S. Hackbart, M.J. Fuenzalida, M.M. Herlihy, H. Ayres, A.R. Dresch, L.M. Vieira, J.N. Guenther, R.R. Grummer, P.M. Fricke, R.D. Shaver, and M.C. Wiltbank. 2014. Relationships between fertility and postpartum changes in body condition and body weight in lactating dairy cows. *J. Dairy Sci.* 97:3666–3671.
- Chapinal, N., S.J. Leblanc, M.E. Carson, K.E. Leslie, S. Godden, M. Capel, J.E. Santos, M.W. Overton, and T.F. Duffield. 2012. Herd-level associations of serum metabolites in the transition period with disease, milk production, and early lactation reproductive performance. *J. Dairy Sci.* 95:5676-5682.
- Contreras, G.A., and L.M. Sordillo. 2011. Lipid mobilization and inflammatory responses during the transition period of dairy cows. *Comparative Immunology, Microbiology Infectious Diseases* 34:281–289.
- Dann, H.M., N.B. Litherland, J.P. Underwood, M. Bionaz, A. D'Angelo, J.W. McFadden, and J.K. Drackley. 2006. Diets during far-off and close-up dry periods affect periparturient metabolism and lactation in multiparous cows. *J. Dairy Sci.* 89:3563-3577.
- Dann, H.M., D.E. Morin, M.R. Murphy, G.A. Bollero, and J.K. Drackley. 2005. Prepartum intake, postpartum induction of ketosis, and periparturient disorders affect the metabolic status of dairy cows. *J. Dairy Sci.* 88:3249-3264.
- Dann, H.M., and B.H. Nelson. 2011. Early lactation diets for dairy cattle – focus on starch. In *Proceedings of the 2011 Cornell Nutrition Conference for Feed Manufacturers*, Syracuse, NY.
- DelCurto, H., G. Wu, and M.C. Satterfield. 2013. Nutrition and reproduction: Links to epigenetics and metabolic syndrome in offspring. *Curr. Opin. Clin. Nutr. Metab. Care.* 16:385–391.
- Diskin, M.G., D.R. Mackey, J.F. Roche, and J.M. Sreenan. 2003. Effects of nutrition and metabolic status on circulating hormones and ovarian follicle development in cattle. *Anim. Reprod. Sci.* 78:345–370.
- Douglas, G.N., T.R. Overton, H.G. Bateman, H.M. Dann, and J.K. Drackley. 2006. Prepartal plane of nutrition, regardless of dietary energy source, affects periparturient metabolism and dry matter intake in Holstein cows. *J. Dairy Sci.* 89:2141-2157.
- Drackley, J.K. 1999. Biology of dairy cows during the transition period: The final frontier? *J. Dairy Sci.* 82:2259-2273.
- Drackley, J.K. and F.C. Cardoso. 2014. Prepartum and postpartum nutritional management to optimize fertility in high-yielding dairy cows in confined TMR systems. *Animal.* 8:S1, 5-14.
- Drackley, J.K., H.M. Dann, G.N. Douglas, N.A. Janovick Guretzky, N.B. Litherland, J.P. Underwood, and J.J. Looor. 2005. Physiological and pathological adaptations in dairy cows that may increase susceptibility to periparturient diseases and disorders. *Italian J. Animal Sci.* 4:323-344.

- Drackley, J.K., R.L. Wallace, D. Graugnard, J. Vasquez, B.F. Richards, and J.J. Looor. 2014. Visceral adipose tissue mass in non-lactating dairy cows fed diets differing in energy density. *J. Dairy Sci.* 97:3420-3430.
- Eckert, J.J., R. Porter, A.J. Watkins, E. Burt, S. Brooks, H.J. Leese, P.G. Humpherson, I.T. Cameron, and T.P. Fleming. 2012. Metabolic induction and early responses of mouse blastocyst developmental programming following maternal low protein diet affecting life-long health. *PLoS One*; 7:e52791.
- Fleming, T.P., M.A. Velazquez, J.J. Eckert. D. Embryos, and D. Barker. 2015. *J. Dev. Orig. Health Dis.* 6:377–383.
- Friggens, N.C. 2003. Body lipid reserves and reproductive cycle: Towards a better understanding. *Livestock Production Sci.* 83:219-236.
- Garnsworthy, P.C. 2007. Body condition score in dairy cows: Targets for production and fertility. In *Recent advances in animal nutrition – 2006* (ed. PC Garnsworthy and J Wiseman), pp. 61-86. Nottingham University Press, Nottingham, UK.
- Garnsworthy, P.C., K.D. Sinclair, and R. Webb. 2008. Integration of physiological mechanisms that influence fertility in dairy cows. *Animal.* 2:1144-1152.
- Garverick, H.A., M.N. Harris, R. Vogel-Bluel, J.D. Sampson, J. Bader, W.R. Lamberson, J.N. Spain, M.C. Lucy, and R.S. Youngquist. 2013. Concentrations of nonesterified fatty acids and glucose in blood of periparturient dairy cows are indicative of pregnancy success at first insemination. *J. Dairy Sci.* 96:181-188.
- Grainger C., G.D. Wilhelms, and A.A. McGowan. 1982. Effect of body condition at calving and level of feeding in early lactation on milk production of dairy cows. *Aust. J. Exp. Agric. Anim. Husb.* 22:9-17.
- Graugnard, D.E., M. Bionaz, E. Trevisi, K.M. Moyes, J.L. Salak-Johnson, R.L. Wallace, J.K. Drackley, G. Bertoni, and J.J. Looor. 2012. Blood immunometabolic indices and polymorphonuclear leukocyte function in peripartum dairy cows are altered by level of dietary energy prepartum. *J. Dairy Sci.* 95:1749-1758.
- Graugnard, D.E., K.M. Moyes, E. Trevisi, M.J. Khan, D. Keisler, J.K. Drackley, G. Bertoni, and J.J. Looor. 2013. Liver lipid content and inflammometabolic indices in peripartal dairy cows are altered in response to prepartal energy intake and postpartal intramammary inflammatory challenge. *J. Dairy Sci.* 96:918-935.
- Grum, D.E., J.K. Drackley, R.S. Younger, D.W. LaCount, and J.J. Veenhuizen. 1996. Nutrition during the dry period and hepatic lipid metabolism of periparturient dairy cows. *J. Dairy Sci.* 79:1850-1864.
- Grummer, R.R. 1995. Impact of changes in organic nutrient metabolism on feeding the transition dairy cow. *J. Anim. Sci.* 73:2820-2833.
- Grummer, R.R., M.C. Wiltbank, P.M. Fricke, R.D. Watters, and N. Silva-Del-Rio. 2010. Management of dry and transition cows to improve energy balance and reproduction. *J. Reprod. Dev.* 56(Suppl):S22–28.

- Hammon D.S., I.M. Evjen, T.R. Dhiman, J.P. Goff, and J.L. Walters. 2006. Neutrophil function and energy status in Holstein cows with uterine health disorders. *Vet. Immunol. Immunopathol.* 13:21-29.
- Holcomb, C.S., H.H. Van Horn, H.H. Head, M.B. Hall, and C.J. Wilcox. 2001. Effects of prepartum dry matter intake and forage percentage on postpartum performance of lactating dairy cows. *J. Dairy Sci.* 84:2051-2058.
- Holtenius, K., S. Agenäs, C. Delavaud, and Y. Chilliard. 2003. Effects of feeding intensity during the dry period. 2. Metabolic and hormonal responses. *J. Dairy Sci.* 86:883-891.
- Huhtanen, P., V. Vanhatalo, and T. Varvikko. 2002. Effects of abomasal infusions of histidine, glucose, and leucine on milk production and plasma metabolites of dairy cows fed grass silage diets. *J. Dairy Sci.* 85:204-216.
- Ingvartsen, K.L., R.J. Dewhurst, and N.C. Friggens. 2003. On the relationship between lactational performance and health: Is it yield or metabolic imbalance that cause production diseases in dairy cattle? A position paper. *Livestock Production Sci.* 83:277-308.
- Ingvartsen, K.L., and K. Moyes. 2013. Nutrition, immune function and health of dairy cattle. *Animal* 7 (Suppl. 1):112-122.
- Janovick, N.A., Y.R. Boisclair, and J.K. Drackley. 2011. Prepartum dietary energy intake affects metabolism and health during the periparturient period in primiparous and multiparous Holstein cows. *J. Dairy Sci.* 94:1385-1400.
- Janovick, N.A., and J.K. Drackley. 2010. Prepartum dietary management of energy intake affects postpartum intake and lactation performance by primiparous and multiparous Holstein cows. *J. Dairy Sci.* 93:3086-3102.
- Ji, P., J.S. Osorio, J.K. Drackley, and J.J. Looor. 2012. Overfeeding a moderate energy diet prepartum does not impair bovine subcutaneous adipose tissue insulin transduction and induces marked changes in periparturient gene network expression. *J. Dairy Sci.* 95:4333-4351.
- Jorritsma, R., T. Wensing, T.A. M. Kruip, T. Vos PLAM, and J.P.T.M. Noordhuizen. 2003. Metabolic changes in early lactation and impaired reproductive performance in dairy cows. *Veterinary Research* 34:11-26.
- Kehrli M.E., J.L. Burton, B.J. Nonnecke, and E.K. Lee. 1999. Effects of stress on leukocyte trafficking and immune responses: Implications for vaccination. *Adv. Vet. Med.* 41:61-81.
- Kim, I.H., and G.H. Suh. 2003. Effect of the amount of body condition loss from the dry to near calving periods on the subsequent body condition change, occurrence of postpartum diseases, metabolic parameters and reproductive performance in Holstein dairy cows. *Theriogenology* 60:1445-1456.
- Komaragiri, M.V., and R.A. Erdman. 1997. Factors affecting body tissue mobilization in early lactation dairy cows. 1. Effect of dietary protein on mobilization of body fat and protein. *J. Dairy Sci.* 80:929-937.
- Kunz, P.L., J.W. Blum, I.C. Hart, J. Bickel, and J. Landis. 1985. Effects of different energy intakes before and after calving on food intake, performance and blood hormones and metabolites in dairy cows. *Animal Production* 40:219-231.
- LeBlanc, S.J. 2010. Assessing the association of the level of milk production with reproductive performance in dairy cattle. *J. Reprod. Dev.* 56(Suppl.), S1-S7.

- Loor, J.J., H.M. Dann, N.A. Janovick-Guretzky, R.E. Everts, R. Oliveira, C.A. Green, N.B. Litherland, S.L. Rodriguez-Zas, H.A. Lewin, and J.K. Drackley. 2006. Plane of nutrition prepartum alters hepatic gene expression and function in dairy cows as assessed by longitudinal transcript and metabolic profiling. *Physiological Genomics* 27:29-41.
- Loor, J.J., R.E. Everts, M. Bionaz, H.M. Dann, D.E. Morin, R. Oliveira, S.L. Rodriguez-Zas, J.K. Drackley, and H.A. Lewin. 2007. Nutrition-induced ketosis alters metabolic and signaling gene networks in liver of periparturient dairy cows. *Physiological Genomics* 32:105-116.
- Mann, S., F.A. Leal Yepes, T.R. Overton, J.J. Wakshlag, A.L. Lock, C.M. Ryan, and D.V. Nydam. 2015. Dry period plane of energy: Effects on feed intake, energy balance, milk production, and composition in transition dairy cows. *J. Dairy Sci.* 98:3366–3382.
- McArt, J.A., D.V. Nydam, and G.R. Oetzel. 2012. Epidemiology of subclinical ketosis in early lactation dairy cattle. *J. Dairy Sci.* 95:5056-5066.
- Moyes, K.M., J.K. Drackley, J.L. Salak-Johnson, D.E. Morin, J.C. Hope, and J.J. Loor. 2009. Dietary-induced negative energy balance has minimal effects on innate immunity during a *Streptococcus uberis* mastitis challenge in dairy cows during mid-lactation. *J. Dairy Sci.* 92:4301-4316.
- Mulligan, F.J., and M.L. Doherty. 2008. Production diseases of the transition cow. *Veterinary Journal* 176:3-9.
- Mulligan, F.J., L. O'Grady, D.A. Rice, and M.L. Doherty. 2006. A herd health approach to dairy cow nutrition and production diseases of the transition cow. *Animal Reproduction Sci.* 96:331-353.
- NRC. 2001. *Nutrient Requirements of Dairy Cattle*. Seventh revised edition. Natl. Acad. Press. Washington, DC.
- Ordway, R.S., S.E. Boucher, N.L. Whitehouse, C.G. Schwab, and B.K. Sloan. 2009. Effects of providing two forms of supplemental methionine to periparturient Holstein dairy cows on feed intake and lactational performance. *J. Dairy Sci.* 92:5154–5166.
- Osorio, J.S., P. Ji, J.K. Drackley, D. Luchini, and J.J. Loor. 2013. Supplemental Smartamine M or MetaSmart during the transition period benefits postpartal cow performance and blood neutrophil function. *J. Dairy Sci.* 96:6248–6263.
- Ospina, P.A., D.V. Nydam, T. Stokol, and T.R. Overton. 2010. Association between the proportion of sampled transition cows with increased nonesterified fatty acids and β -hydroxybutyrate and disease incidence, pregnancy rate, and milk production at the herd level. *J. Dairy Sci.* 93:3595-3601.
- Penagaricano, F., A.H. Souza, P.D. Carvalho, A.M. Driver, R. Gamba, J. Kropp, K.S. Hackbart, D. Luchini, R.D. Shaver, M.C. Wiltbank, and H. Khatib. 2013. Effect of maternal methionine supplementation on the transcriptome of bovine preimplantation embryos. *PLoS One.* 8:e72302.
- Pisulewski, P.M., H. Rulquin, J.L. Peyraud, and R. Verite. 1996. Lactational and systemic responses of dairy cows to postruminal infusions of increasing amounts of methionine. *J. Dairy Sci.* 79:1781–1791.
- Robinson, J.J., C.J. Ashworth, J.A. Rooke, L. M. Mitchell, and T. G. McEvoy. 2006. Nutrition and fertility in ruminant livestock. *Anim. Feed Sci. Technol.* 126:259–276.

- Roche, J.R., A.W. Bell, T.R. Overton, and J.J. Looor. 2013. Nutritional management of the transition cow in the 21st century – a paradigm shift in thinking. *Animal Production Sci.* 53:1000-1023.
- Rukkwamsuk, T, T. Wensing, and M.J. Geelen. 1998. Effect of overfeeding during the dry period on regulation of adipose tissue metabolism in dairy cows during the periparturient period. *J. Dairy Sci.* 81:2904-2911.
- Rukkwamsuk, T, T. Wensing, and T.A.M. Kruip. 1999. Relationship between triacylglycerol concentration in the liver and first ovulation in postpartum dairy cows. *Theriogenology.* 51:1133-1142.
- Santos, J.E., R.S. Bisinotto, E.S. Ribeiro, F.S. Lima, L.F. Greco, C.R. Staples, and W.W. Thatcher. 2010. Applying nutrition and physiology to improve reproduction in dairy cattle. *Soc. Reprod. Fertil. Suppl.* 67:387–403.
- Santos, J.E., R.L. Cerri, and R. Sartori. 2008. Nutritional management of the donor cow. *Theriogenology.* 69:88–97.
- Santos, J.E., E.J. DePeters, P.W. Jardon, and J.T. Huber. 2001. Effect of prepartum dietary protein level on performance of primigravid and multiparous Holstein dairy cows. *J. Dairy Sci.* 84:213–224.
- Santos, J.E., H.M. Rutigliano, and M.F. Sa Filho. 2009. Risk factors for resumption of postpartum estrous cycles and embryonic survival in lactating dairy cows. *Animal Reproduction Sci.* 110:207-221.
- Silva-del-Rio, N, P.M. Fricke, and R.R. Grummer. 2010. Effects of twin pregnancy and dry period strategy on milk production, energy balance, and metabolic profile in dairy cows. *J. Animal Sci.* 88:1048-1060.
- Silvestre, F.T., T.S.M. Carvalho, N. Francisco, J.E.P. Santos, C.R. Staples, T.C. Jenkins, and W.W. Thatcher. 2011. Effects of differential supplementation of fatty acids during the peripartum and breeding periods of Holstein cows: I. Uterine and metabolic responses, reproduction, and lactation. *J. Dairy Sci.* 94:189-204.
- Sinclair, K.D., C. Allegrucci, R. Singh, D.S. Gardner, S. Sebastian, J. Bispham, A. Thurston, J.F. Huntley, W.D. Rees, C.A. Maloney, R.G. Lea, J. Craigon, T.G. McEvoy, and L.E. Young. 2007. DNA methylation, insulin resistance, and blood pressure in offspring determined by maternal periconceptional B vitamin and methionine status. *Proc. Natl. Acad. Sci U S A.* 104: 19351–19356.
- Stella, S.L. 2017. Uterine immune mediators and lipid profiles of the preimplantation embryo and endometrial tissue in Holstein cows supplemented with rumen-protected methionine and observed neutrophil extracellular trap (net) formation in the bovine endometrial tissue. MS Thesis. University of Illinois, Urbana.
- Sutter, F., and D.E. Beever. 2000. Energy and nitrogen metabolism in Holstein-Friesen cows during early lactation. *Animal Sci.* 70:503-514.
- Thatcher, W.W., J.E.P. Santos, and C.R. Staples. 2011. Dietary manipulations to improve embryonic survival in cattle. *Theriogenology* 76:1619-1631.
- Toledo, M., G.M. Baez, E. Trevisol, N.E. Lobos, A. Garcia-Guerra, J.N. Guen, D. Luchini, R.D. Shaver, and M.C. Wiltbank. 2015. Effect of top-dressing rumen-protected methionine in lactating Holstein cows II: Fertility and embryo development. *J. Dairy Sci.* V. 98 (Suppl. 2):301.

- Vickers, L.A., D.M. Weary, D.M. Veira, and M.A.G. von Keyserlingk. 2013. Feeding a higher forage diet prepartum decreases incidences of subclinical ketosis in transition dairy cows. *J. Animal Sci.* 91:886-894.
- Walsh, R.B., J.S. Walton, D.F. Kelton, S.J. LeBlanc, K.E. Leslie, and T.F. Duffield. 2007. The effect of subclinical ketosis in early lactation on reproductive performance of postpartum dairy cows. *J. Dairy Sci.* 90:2788-2796.
- Wang, J, Z. Wu, D. Li, N. Li, S.V. Dindot, M.C. Satterfield, F.W. Bazer, and G. Wu. 2012. Nutrition, epigenetics, and metabolic syndrome. *Antioxid Redox Signal.* 17:282–301.
- Whitaker, D.A., E.J. Smith, G.O. da Rosa, and J.M. Kelly. 1993. Some effects of nutrition and management on the fertility of dairy cattle. *Veterinary Record* 133:61-64.
- Wiltbank, M.C., H. Lopez, R. Sartori, S. Sangsritavong, and A. Gumen. 2006. Changes in reproductive physiology of lactating dairy cows due to elevated steroid metabolism. *Theriogenology.* 65:17–29.
- Winkleman, L.A., T.H. Elsasser, and C.K. Reynolds. 2008. Limit-feeding a high-energy diet to meet energy requirements in the dry period alters plasma metabolite concentrations but does not affect intake or milk production in early lactation. *J. Dairy Sci.* 91:1067-1079.
- Wu, G., F.W. Bazer, M.C. Satterfield, X. Li, X. Wang, G.A. Johnson, R.C. Burghardt, Z. Dai, J. Wang, and Z. Wu. 2013. Impacts of arginine nutrition on embryonic and fetal development in mammals. *Amino Acids* 45:241-256.
- Zabeli, Q., and B.U. Metzler-Zabeli. 2012. Interplay between rumen digestive disorders and diet-induced inflammation. *Research in Veterinary Sci.* 93:1099-1108.
- Zerbe, H., N. Schneider, W. Leibold, T. Wensing, T.A. Kruij, and H.J. Schuberth. 2000. Altered functional and immunophenotypical properties of neutrophilic granulocytes in postpartum cows associated with fatty liver. *Theriogenology* 54:771-86.

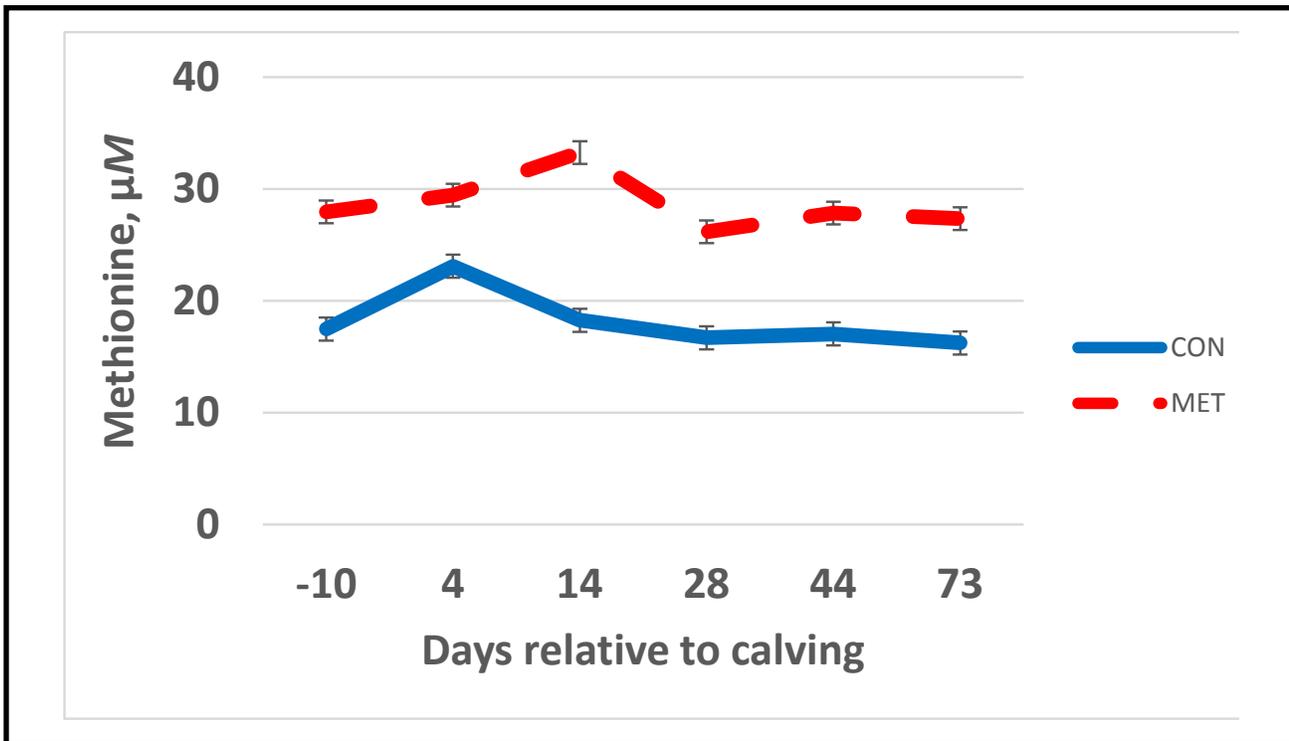


Figure 1. Serum methionine concentration was greater ($P < 0.05$) in cows fed rumen-protected methionine (MET; $n = 10$) than cows not fed rumen-protected methionine (CON; $n = 7$)(Stella, 2017).